



Paresordin A, a new diphenyl cyclic peroxide from the lichen *Parmotrema praesorediosum*

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ABSTRACT

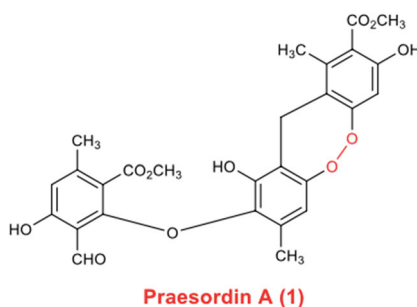
From the lichen *Parmotrema praesorediosum*, one new diphenyl peroxide, named praesordin A (1), together with four depsidones, including virensic acid (2), protocetraric acid (3), 8'-O-methylprotocetraric acid (4), and furfuric acid (5) were purified. Their structures were characterized using extensive HR-ESI-MS and NMR spectroscopic methods. The isolated compounds (2–5) possessed stronger α -glucosidase inhibitory activity ($IC_{50} = 43.7$ – $110.1 \mu\text{M}$) than the standard drug acarbose ($IC_{50} = 214.5 \mu\text{M}$).

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1. Introduction

The *Parmotrema* (Parmeliaceae) is a large genus with approximately 350 species of foliose lichens and a high level of diversity in the tropical areas of the world. Phytochemical studies evidenced γ -lactone acids as the main component of *P. praesorediosum* [1–3]. Further, our previous papers evinced diphenyl ethers were

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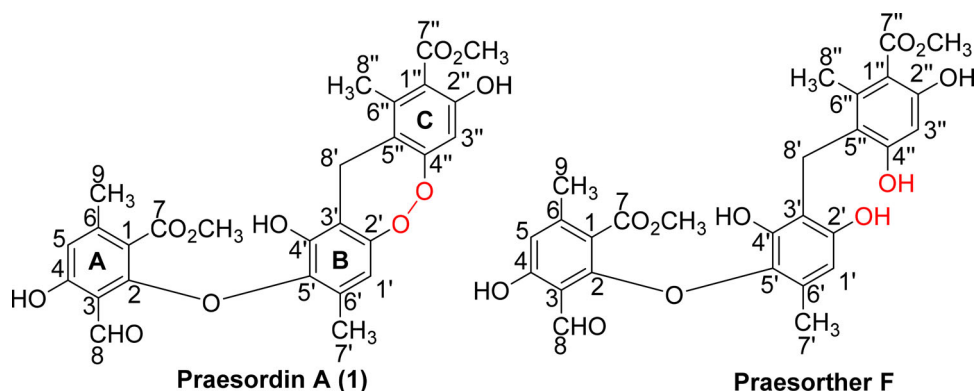


Figure 1. Chemical structures of compound 1 and praesorether F.

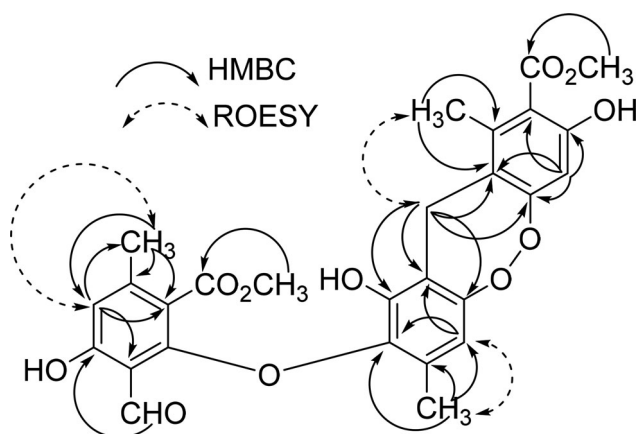
detected in this species at high concentrations [4–6]. As part of our continuing investigation of α -glucosidase inhibitors [7–9], this paper details the separation, structural characterization, and α -glucosidase inhibition of one new diphenyl cyclic peroxide, named praesordin A (1), together with four depsidones, including virensic acid (2), protocetraric acid (3), 8'-*O*-methylprotocetraric acid (4) [10], and furfural (5) [11] from *P. praesorediosum* collected in Dong Nai province, Vietnam (Figure 1).

2. Results and discussion

Compound 1 was isolated as a yellow solid. Its molecular formula was established as $C_{27}H_{24}O_{11}$ by HR-ESI-MS data (m/z 547.1212 $[M + Na]^+$; m/z 525.1396 $[M + H]^+$). IR absorptions detailed the presence of the hydroxyl (3394 cm^{-1}), carbonyl (1708 cm^{-1}), and olefin (1647 cm^{-1}) moieties. The $^1\text{H-NMR}$ spectrum of 1 (Table 1) exhibited one aldehyde proton at δ_{H} 10.44 (1H, s, H-8), three aromatic protons at δ_{H} 6.48 (1H, *dd*, 0.5 & 0.5 Hz, H-5), 6.18 (1H, *d*, 0.5 Hz, H-1'), 6.31 (1H, *s*, H-3''), two methoxy signals at δ_{H} 3.17 (3H, *s*, 7-OCH₃) and 3.86 (3H, *s*, 7''-OCH₃), two methylene protons at δ_{H} 3.97 (1H, *brs*, H-8'a) and 3.98 (1H, *brs*, H-8'b), and three methyl groups at δ_{H} 2.13 (3H, *d*, 0.5 Hz, H-9), 1.98 (3H, *s*, H-7'), 2.61 (3H, *s*, H-8''). The combination of $^{13}\text{C-NMR}$ and HSQC spectra of 1 (Table 1) revealed 27 carbons comprising one formyl carbon at δ_{C} 195.6 (C-8), two carbonyl carbons at δ_{C} 166.8 (C-7), 172.8 (C-7''), eighteen aromatic carbons in the range of 102.0–164.2 ppm (seven oxygenated and four methine carbons), two methoxy carbons at δ_{C} 52.0 (7-OCH₃), 51.9 (7''-OCH₃), one methylene carbon at δ_{C} 21.1 (C-8'), and three methyl carbons at δ_{C} 20.5 (C-9), 16.7 (C-7'), 19.5 (C-8''), which were proved the diphenyl ether skeleton owned three aromatic rings, in which rings A and B linked through an ether bridge, while rings B and C connected together via a methylene linkage at its C-3' and C-5', similar to praesorether F [6]. However, the molecular formula $C_{27}H_{24}O_{11}$ of 1 affirmed sixteen degrees of unsaturation, supported by HR-ESI-MS data. Three aromatic rings, one formyl group, and two carbonyl moieties were accounted for fifteen degrees of unsaturation in 1 [6], and one existing degree of unsaturation was secured the present of the peroxide ring in 1. On the other hands, the HMBC spectrum of 1 (Figure 2) signified correlations between two methylene protons at δ_{H} 3.97 (H-8'a),

Table 1. ^1H and ^{13}C NMR spectral data for compound **1** and praesorether F in acetone- d_6 .

Position	1		Praesorether F	
	δ_{H}	δ_{C}	δ_{H}	δ_{C}
1		115.7		115.5
2		159.2		158.9
3		111.0		111.0
4		164.2		164.3
5	6.48 (<i>dd</i> , 0.5 & 0.5)	112.7	6.52 (<i>d</i> , 0.5)	112.9
6		148.1		148.2
7		166.8		166.6
8	10.44 (<i>s</i>)	195.6	10.44 (<i>s</i>)	195.6
9	2.13 (<i>d</i> , 0.5)	20.5	2.15 (<i>d</i> , 0.5)	20.5
1'	6.18 (<i>d</i> , 0.5)	108.5	6.24 (<i>s</i>)	108.9
2'		153.9		153.3
3'		113.7		113.1
4'		149.8		149.2
5'		135.7		135.8
6'		129.6		129.9
7'	1.98 (<i>s</i>)	16.7	1.99 (<i>s</i>)	16.6
8'	3.97 (<i>brs</i>)	21.1	3.97 (<i>brs</i>)	20.7
	3.98 (<i>brs</i>)		3.98 (<i>brs</i>)	
1''		107.1		109.4
2''		162.3		161.5
3''	6.31 (<i>s</i>)	102.0	6.33 (<i>s</i>)	101.5
4''		163.5		160.0
5''		120.8		119.5
6''		142.1		142.5
7''		172.8		172.4
8''	2.61	19.5	2.58	19.1
7-OCH ₃	3.17	52.0	3.18	52.0
7''-OCH ₃	3.86	51.9	3.88	52.1

**Figure 2.** Key HMBC and ROESY correlations of **1**.

3.98 (H-8'b) and two oxygenated aromatic carbons at δ_{C} 153.9 (C-2'), 163.5 (C-4''), and furthermore, the observed downfield shifts of these carbons [at δ_{C} 153.9 (C-2'), 163.5 (C-4'') of **1**, and δ_{C} 153.3 (C-2'), 160.0 (C-4'') in praesorether F, in acetone- d_6] (Table 1), which were designated the 2',4''-peroxy bridge. According to the above-mentioned analysis, the structure of **1** was established as shown, named praesordin A.

Table 2. α -Glucosidase inhibition of compounds 2–5.

Samples	Inhibition (%)					IC ₅₀ (μ M)
	250	100	50	25	10	
2	94.50 \pm 1.10	46.79 \pm 0.75	19.35 \pm 0.69	2.40 \pm 1.00	–	110.1
3	*	68.47 \pm 0.95	23.90 \pm 1.40	12.41 \pm 0.84	6.70 \pm 1.20	79.3
4	*	75.68 \pm 0.88	51.94 \pm 0.91	25.20 \pm 1.80	4.70 \pm 1.20	48.2
5	*	92.80 \pm 1.30	56.90 \pm 1.30	29.3 \pm 1.10	10.30 \pm 1.20	43.7
Acarbose						214.5

*Inhibition > 100%.

- Inhibition < 1%.

Compounds (2-5) were measured for their α -glucosidase inhibitory activity (Table 2). As results, compounds (2-5) expressed more potent inhibition against enzyme α -glucosidase (IC₅₀ ranged from 43.7 to 110.1 μ M) than the standard drug acarbose (IC₅₀ = 214.5 μ M).

Some natural cyclic peroxides were announced from others origins [12–14], however, this is the first time that diphenyl cyclic peroxide was enunciated from the lichen *Parmotrema praesorediosum*.

3. Experimental

3.1. General experimental procedures

The IR spectra were measured on Shimadzu FTIR-8200 infrared spectrophotometer. The HR-ESI-MS were recorded on a HR-ESI-MS MicroOTOF-Q mass spectrometer or on a LC-Agilent 1100 LC-MSD Trap spectrometer. NMR spectra were recorded on a Bruker Avance III spectrometer, using residual solvent signal as internal reference: acetone-*d*₆ δ _H 2.05, δ _C 206.31 and 30.6, at 500 MHz for ¹H NMR and 125 MHz for ¹³C NMR. TLC was carried out on precoated silica gel 60 F₂₅₄ or silica gel GF₂₅₄ (Merck). Spots were visualized by spraying with 10% aqueous H₂SO₄ or 5% ferric chloride solutions followed by heating. Gravity column chromatography was performed with silica gel 60 (0.040–0.063 mm, Himedia).

3.2. Plant material

The lichen thalli of *Parmotrema praesorediosum* were collected on the bark of *Dipterocarpus* sp. at Tan Phu forest, Dong Nai province, Vietnam in June 2009. The geographical location where the lichen was collected is at an altitude of 110 m, 11°20'–11°50' N and 107°09'–107°35' E. The botanical species of *Parmotrema praesorediosum* (Nyl.) Hale (synonym of *Parmelia praesorediosa* Nyl.) was identified by Dr. Vo Thi Phi Giao, Faculty of Biology, University of Science, National University – Ho Chi Minh City. A voucher specimen (No US-B020) was deposited in the Herbarium of The Department of Organic Chemistry, Faculty of Chemistry, University of Science, National University – Ho Chi Minh City, Vietnam.

3.3. Extraction and isolation

The thallus material (5.0 kg) was washed under flow of tap water and then was air-dried at ambient temp. to obviate thermally induced decomposition prior to being ground into a fine powder. The ground powder sample (3.0 kg) was macerated by methanol at room temperature to afford a crude methanol extract (450 g). This crude one (450 g) was applied to silica gel solid phase extraction, successively eluted with the following solvents: petroleum ether (60–90 °C) (PE), chloroform (C), ethyl acetate (EA), acetone (A) and methanol (M) to afford corresponding extracts: extract PE (40 g), extract C (105 g), extract EA (50 g), extract A (45 g) and extract M (37 g).

The chloroform extract (105 g) was subjected to silica gel column chromatography, eluted by the solvent system of petroleum ether – ethyl acetate with increasing ethyl acetate to give twenty three fractions, coded C1–C23. Fraction C16 (4.2 g) was rechromatographed, eluted with chloroform-methanol (95:5) to give **5** (15.0 mg). Fraction C19 (6.1 g) was applied on silica gel column and eluted with a gradient solvent system of chloroform-acetone (95:5) to give three fractions (C19a, C19b and C19c). Fraction C19b (3.2 g) was rechromatographed, eluted with chloroform-acetone (98:2) to give six fractions (C19ba to C19bf). Fraction C19ba (169.6 mg) was subjected to pre TLC (chloroform-methanol, 95:5, 9:1 and *n*-hexane-diethyl ether, 5:5) to afford compound **1** (10.5 mg).

The acetone extract (45.0 g) was applied to silica gel column chromatography and eluted with ethyl acetate–methanol (9:1-5:5) to give 6 fractions (from AC.1 to AC.6). Subfraction AC2 (2.5 g) was applied to column chromatography, eluting with chloroform–acetone–acetic acid (95:5:3 drops) to yield **2** (5.2 mg) and **4** (5.0 mg). Fraction AC5 (28 g) was rechromatographed, eluted with chloroform-acetone (7:3-5:5) to give **3** (2.0 g).

3.3.1. Praesordin A (1)

Yellow solid; IR (KBr) ν_{\max} : 3394, 1708, 1647, 1272 cm^{-1} . $^1\text{H-NMR}$ (acetone- d_6 , 500 MHz, *J* in Hz) and $^{13}\text{C-NMR}$ (acetone- d_6 , 125 MHz) spectral data see [Table 1](#); HR-ESI-MS: *m/z* 547.1212 [$\text{M} + \text{Na}$] $^+$ (calcd for $\text{C}_{27}\text{H}_{24}\text{O}_{11}\text{Na}$, 547.1216); 525.1396 [$\text{M} + \text{H}$] $^+$ (calcd for $\text{C}_{27}\text{H}_{25}\text{O}_{11}$, 525.1397).

3.4. α -Glucosidase inhibition assay

Compounds **2-5** were evaluated their inhibitory activity against enzyme α -glucosidase as previously described paper, and acarbose was used as the positive control [7–9].

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Disclosure statement

No potential conflict of interest was reported by the authors.

References

- [1] F. David, J.A. Elix, and M.W.D. Samsudin, *Aust. J. Chem.* **43**, 1297 (1990).
- [2] U. Jayalal, P.K. Divakar, S. Joshi, S.O. Oh, Y.J. Koh, and J.S. Hur, *Mycobiology* **41**, 25 (2013).
- [3] B.L.C. Huynh, T.H. Duong, T.M.L. Do, T.G. Pinnock, L.M. Pratt, S. Yamamoto, H. Watarai, T. Tanahashi, and K.P.P. Nguyen, *Rec. Nat. Prod.* **10**, 332 (2016).
- [4] B.L.C. Huynh, T.H. Duong, T. Tanahashi, and K.P.P. Nguyen, *Vietnam J. Chem.* **48**, 332 (2010).
- [5] B.L.C. Huynh, H.D. Le, Y. Takenaka, T. Tanahashi, and K.P.P. Nguyen, *Magn. Reson. Chem.* **54**, 81 (2016).
- [6] B.L.C. Huynh, V.M. Bui, K.P.P. Nguyen, N.K.T. Pham, and T.P. Nguyen, *Nat. Prod. Res.* (2020).
- [7] T.P. Nguyen, T.T.V. Tran, D.T. Mai, T.D. Le, N.M. Phan, and T.D. Bui, *Nat. Prod. Res.* **29**, 1432 (2015).
- [8] T.P. Nguyen, T.D. Le, N.M. Phan, T.D. Bui, and D.T. Mai, *J. Asian Nat. Prod. Res.* **18**, 542 (2016).
- [9] T.P. Nguyen, T.D. Le, N.M. Phan, T.D. Bui, N.K.T. Pham, T.M.L. Do, D.T. Nguyen, and D.T. Mai, *Nat. Prod. Res.* **30**, 2389 (2016).
- [10] V.K. Nguyen, and T.H. Duong, *Vietnam J. Sci. Technol. Eng.* **61**, 12 (2019).
- [11] J. Gunzinger, and R. Tabacchi, *Helv. Chim. Acta* **68**, 1936 (1985).
- [12] D.Q. Yu, R.Y. Chen, L.J. Huang, F.Z. Xie, D.S. Ming, K. Zhou, H.Y. Li, and K.M. Tong, *J. Asian Nat. Prod. Res.* **10**, 851 (2008).
- [13] P. Jumaryatno, L.K. Lambert, J.N.A. Hooper, J.T. Blanchfield, and M.J. Garson, *Nat. Prod. Commun.* **8**, 725 (2013).
- [14] V.M. Dembitsky, *J. Mol. Genet. Med.* **9**, 1000163 (2015).